# Taxonomy and distribution of freshwater *Prasiola* (Prasiolales, Chlorophyta) in central Mexico

Rocío RAMÍREZ Rodríguez & Javier CARMONA Jiménez\*

Departamento de Ecología y Recursos Naturales, UNAM. A.P. 70-620, Ciudad Universitaria, Coyoacán, C.P. 04510, México, D.F.

(Received 2 September 2003, accepted 23 April 2004)

**Abstract** – Seventeen samples and specimens of *Prasiola* from central Mexico, representing two species, have been studied in laboratory culture. The most common species, *P. mexicana*, is characterized by large leaf-like thalli ( $\geq$  90 mm), bifurcating branches and a smooth margin. The lamina apex has numerous aplanospores and the margin is polystromatic ( $\geq$  8 layers). Specimens with a corrugated surface and rhizoidal cells in the middle region were also noted. The second species, *P. nevadensis*, is characterized by small, leaf-like thalli ( $\geq$  30 mm), crenate margins, and a monostromatic lamina. Two relevant morphological features were noted in culture: multilocular gametangia with possible female gametes, and the formation of small cellular packets fixed to the apical margin of the lamina. *Prasiola nevadensis* is here reported for the first time for Mexico, which also represents only the second report for the species. *Prasiola mexicana* was found in subaerial or submerged conditions in cold rivers (2-16.4°C), neutral pH (6.4-7.6) and low specific conductivity (35.5-94.2 μS.cm<sup>-1</sup>). *Prasiola nevadensis* was collected submerged in a river with cold water (14°C), acidic pH (5.5), and moderate specific conductivity (192 μS·cm<sup>-1</sup>). Both species were collected in high mountain regions ( $\geq$  1300 masl).

Chlorophyta / distribution / México / Prasiola / Prasiolales / streams / taxonomy

Résumé – Taxinomie et distribution de *Prasiola* (Prasiolales, Chlorophyta) dulcaquicoles au Mexique central. On a observé deux espèces parmi dix-sept récoltes et spécimens de Prasiola du Mexique central maintenus en culture. La plus commune, P. mexicana, est caractérisée par de larges thalles foliacés (≥ 90 mm), des ramifications bifurquées et une marge lisse; le sommet des thalles porte de nombreux aplanospores et montre une marge polystromatique (≥ 8 couches). On remarque aussi des spécimens avec une surface plissée et des cellules rhizoïdales dans la région médiane. La seconde espèce, P. nevadensis, est caractérisée by de petits thalles foliacés (≥ 30 mm), une marge crénelée et une lame monostromatique. Deux caractères morphologiques pertinents ont été notés en culture : des gamétanges multiloculaires contenant d'éventuels gamètes femelles et la formation de petits paquets cellulaires fixés à la marge apicale du thalle. Prasiola nevadensis, connu uniquement de son lieu d'origine, est signalé ici pour la première fois du Mexique. Prasiola mexicana a été trouvé en conditions subaériennes ou submergées dans des rivières froides (2-16.4 °C), à pH neutre (6.4-7.6) et à conductivité spécifique basse (35.5-94.2 μS.cm<sup>-1</sup>). Prasiola nevadensis a été récolté submergé dans une rivière à eau fraîche (14 °C), à pH acide (5.5) et à conductivité spécifique modérée (192 µS.cm<sup>-1</sup>). Les deux espèces ont été récoltées dans des régions de haute montagne (altitude ≥ 1 300 m).

Chlorophyta / distribution / México / Prasiola / Prasiolales / rivières / taxonomie

<sup>\*</sup> Correspondence and reprints: jcj@hp.fciencias.unam.mx Communicating editor : Pierre Compère

#### INTRODUCTION

The genus *Prasiola* is included in the class Trebouxiophyceae and is defined by the following characteristics: a monostromatic laminar thallus, vegetative cells with stellate or lobed chloroplasts containing a single pyrenoid, biflagellate male gametes with cross-type flagellar insertion and a basal body disposed in counterclockwise fashion, closed mitosis with a persistent spindle in telophase, and cytokinesis by transverse wall deposition (O'Kelly et al., 1989; Sherwood et al., 2000). The systematic position of the Prasiolales has been controversial due to the similarity of the type of mitosis and cytokinesis to the class Ulvophyceae. However, Friedl & O'Kelly (2002) analyzed the 18S rDNA nucleotide sequence and confirmed its placement in the Trebouxiophyceae. The family Prasiolaceae includes two genera: Rosenvingiella P.C. Silva, characterized by its filamentous organization, and *Prasiola* G. Meneghini, which has a monostromatic laminar thallus and oogametic reproduction (Friedmann, 1959a, 1959b; Bravo, 1965; Rindi et al., 1999). The delimitation of these two genera has been debated since some species of Prasiola have filamentous stages similar to Rosenvingiella (Bravo, 1965). Recently, Sherwood et al. (2000) found molecular differences between the two genera based on analysis of rbcL and 18S rRNA nucleotide sequences.

Species of the genus *Prasiola* are common inhabitants of marine, freshwater and terrestrial environments in temperate and tropical boreal regions (Naw & Hara, 2002). There are nine currently recognized freshwater species of *Prasiola* (Starmach, 1972; Naw & Hara, 2002): five reported in Asia, one with two varieties (*P. japonica* Yatabe, *P. formosana* Okada var. *formosana*, *P. formosana* Okada var. *coreana*, *P. sinica* Jao, *P. subareolata* Skuja, *P. yunnanica* Jao) and four reported in North America and Europe (*P. fluviatilis* (Sommerfeld) Areschoug, *P. nevadensis* Setchell *et* Gardner, *P. velutina* (Lyngbye) Wille and *P. mexicana* J. Agardh). The characteristics used to differentiate species include the habit, size, and thickness of the lamina, and the disposition, length, and cell size of the foot used to attach to the substratum (Starmach, 1972). Even so, identification is often problematic due to the subjectivity of observers and variability of some characters.

One of the species of the order Prasiolales, *P. mexicana*, has been found in six waterways of the high mountains of central Mexico (Ortega, 1984; Ramírez *et al.*, 2001). However, information about its anatomical characteristics (vegetative and reproductive) and distribution is lacking. In this work, the anatomical structure, habitat, and geographic distribution of populations of *Prasiola* are presented in detail in addition to previous reports from the central region of Mexico.

## **MATERIALS AND METHODS**

A total of 17 samples of *Prasiola* were collected from 8 localities between 19°-20° N, 98°-99° W, and at altitudes of 1,330 to 3,220 m. (Fig. 1). Samples were preserved in 3% formalin and deposited in **FCME** (Holmgren *et al.*, 1990). The physical and chemical characteristics of the water were measured with a Conductronic electro-chemical analyzer (pH  $\pm$  0.02; conductivity  $\pm$  0.5 %; temperature  $\pm$  0.5).

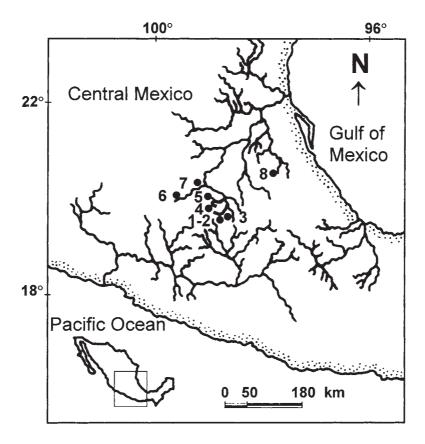


Fig. 1. Location of the study region in central Mexico with freshwater populations of *Prasiola* indicated (circles).

All of the characters considered to be of taxonomic importance in *Prasiola* were examined (Agardh, 1847; Collins 1909; Setchell & Gardner, 1920; Friedmann, 1959a, 1959b; Bravo, 1965; Starmach, 1972; Hamilton & Edlund, 1994; Van den Hoek *et al.*, 1995; Naw & Hara, 2002). Measurements were made in replicates of 20 and are represented as an interval containing a maximum, minimum, average, and standard deviation. The number of replicates was determined using the equation:  $n = (S/E\overline{x})^2$ , where S = standard deviation,  $\overline{x} = \text{average}$  and E = a predetermined standard error (Southwood, 1978). Each specimen was observed and measured with a Olympus BX51 contrast interference microscope, with an SC35 system of microphotography. The thickness of the lamina was examined in sections made with an ultrafreeze microtome (Sartorius-Werke Mod. 27, Germany). A Student's t-test was used to compare morphological characters (gametes, aplanospores and vegetative cells).

Reproductive structures were observed in specimens from two populations maintained in culture: Xalancocotla, *coll. R. Ramírez*, 14.i.2003 and Malila, *coll. J. Carmona*, 01.iii.2003. Cultures were initiated from 10-15 mm segments of

thallus placed in sterilized tubes with distilled water and then vigorously agitated to remove epiphytes. The segments were later rinsed three times in distilled water in Petri dishes, and placed in culture tubes with 50 ml of culture media. Three replicates were transferred from each population. Each specimen was maintained in a 20:1 water-soil culture media for 60 days in climate-controlled chambers with fluorescent lights (Phillips 15 W) under the following conditions: temperature 14°C; irradiance 98-150  $\mu$ mol photons m $^{-2}$  s $^{-1}$  and a photoperiod of 12 h. A solution of Iodine (Johansen, 1940) was used to stain the pyrenoids.

## RESULTS AND DISCUSSION

## **Morphological Analysis**

Two groups of populations were recognized in central Mexico, both with characteristics referable to the genus *Prasiola* (Table 1, Figs 2-20): a laminar monostromatic thallus with a central chloroplast with lobed margin, a central pyrenoid surrounded by plates of starch, and oval to square vegetative cells generally arranged in packets of four (Figs 2-3, 6, 11-12, 15).

The first group (Table 1, Figs 2-10) included the 14 populations from the watershed of Mexico and corresponds to the descriptions of P. mexicana by Agardh (1847), Collins (1909) and Setchell & Gardner (1920) (Table 2). It was characterized by large ( $\geq 9.0$  cm long), procumbent lamina with oval to irregular shape (Fig. 2) with one or many  $(\geq 4)$  fronds originating from the same base and a smooth lamina margin (Fig. 4). Bifurcating ramifications were sometimes present in the apical part of the lamina. It attaches to the substratum with a small mass of elongated, fibrous cells that form a rhizoidal holdfast (Fig. 5). The basal and intermediate sections of the lamina were monostromatic (Fig. 6), while the apical region had abundant aplanospores (≥ 8 layers) and a tendency to become polystromatic (≥ 130 µm thick) (Fig. 7). Vegetative cells were oval to square, 3.1-16.0 µm in diameter (average 9.2 µm); length 6.1-19.8 µm (average 12.2 µm). Aplanospores were seen in both field and cultured specimen, were square or rounded, with a granulate cellular content, a diameter of 5.4-10.6 µm (average 7.7 µm), and a length of 4.5-10.8 µm (average 7.9 µm) (Fig. 7); these were easily distinguished superficially from the vegetative cells, and were found isolated or in groups (Figs 3, 5) in either the basal or apical region of the lamina. The aplanospores were liberated from the apical margin of the lamina upon rupture of the cell wall.

Two interesting morphological characteristics were observed in several specimens (populations 2-8). The surface of the lamina was frequently corrugated (Fig. 9) in specimens collected from terrestrial and high-velocity currents (Table 2), which seems to be related to the physical factors (current velocity or substratum), or biotic interactions (fungal or larval infections). This feature has not been described in any other freshwater species and further studies to characterize the microhabitat and morphological variation will be of interest. The second novel character was the presence of elongated and colorless cells in the folds of the corrugated specimen (Fig. 10). These cells are similar to those described by Fritsch (1961) and Naw & Hara (2002) as rhizoidal cells.

Table 1. Characteristics of freshwater populations of *Prasiola* in Mexico. Measurements are in µm and represent ranges, average and standard deviation.

| Population                         | Lamina                      | Lamina                     | LL/LD                     | Thickness                   | Veget                      | ative cell                | Aplano                   | sporangia               |
|------------------------------------|-----------------------------|----------------------------|---------------------------|-----------------------------|----------------------------|---------------------------|--------------------------|-------------------------|
| and<br>locality                    | length<br>(mm)<br>LL        | diameter<br>(mm)<br>LD     |                           |                             | Length                     | Diameter                  | Length                   | Diameter                |
| 1. MA87<br>Trout tank              | 7.0-10.0<br>8.8 ± 1.3       | 5.0-9.0<br>6.4 ± 1.6       | 1.1-1.8<br>1.4 ± 0.2      | 24.6-32.9<br>28.8 ± 2.9     | 8.3-16.0<br>11.2 ± 2.3     | 6.3-16.0<br>8.4 ± 1.8     | 4.5-8.8<br>6.9 ± 1.4     | 5.4-10.6<br>8.1 ± 2.2   |
| 2. MA1<br>Xalancocotla             | 5.0-26.0<br>14.4 ± 8.4      | 3.0-14.0<br>$8.4 \pm 4.7$  | 0.7-2.6<br>$1.8 \pm 0.7$  | 22.3-27.5<br>$24.9 \pm 1.8$ | 6.1-18.5<br>10.7 ± 2.7     | 3.1-13.5<br>$7.1 \pm 2.4$ | -<br>-                   | _<br>_                  |
| 3. MA9<br>Xalancocotla             | 2.0-14.0<br>$8.3 \pm 6.0$   | 6.0-19.0<br>$14.3 \pm 7.2$ | 0.7-1.5<br>$1.1 \pm 0.3$  | -                           | 9.7-15.5<br>12.1 ± 1.8     | 7.2-11.7<br>$8.9 \pm 1.3$ | -                        | -                       |
| 4. MA26<br>Xalancocotla            | 10.0-18.0<br>14.6 ± 3.2     | 4.0-21.0<br>10.0 ± 6.5     | 0.6-3.7<br>1.9 ± 1.1      | 69.0-130.0<br>88.3 ± 16.3   | 7.0-14.6<br>11.0 ± 2.1     | 3.6-9.4<br>7.6 ± 1.6      | -                        | -                       |
| 5. MA36<br>Xalancocotla            | 4.0-17.0<br>9.5 ± 5.5       | 4.0-15.0<br>9.0 ± 4.9      | $0.2-1.7$ $1.3 \pm 0.7$   | _                           | $7.2-18.3$ $12.4 \pm 2.3$  | 7.2-10.3<br>9.1 ± 0.9     | _                        | _                       |
| 6. MA102<br>Xalancocotla           | 5.0-6.0<br>$5.5 \pm 0.7$    | $4.0-5.0$ $4.5 \pm 0.7$    | $1.0-1.5$ $1.2 \pm 0.3$   | _                           | 9.4-19.8<br>13.7 ± 3.5     | 7.6-12.8<br>10.3 ± 1.8    | -                        | _                       |
| 7. MA94<br>Purification            | $6.0-15.0$ $10.0 \pm 3.7$   | 9.0-29.0<br>15.0 ± 8.4     | $0.3-1.6$ $0.7 \pm 0.5$   | -<br>-                      | 9.4-15.8<br>11.9 ± 1.9     | 8.1-11.5<br>9.3 ± 0.9     | -<br>-                   | -<br>-                  |
| 8. MA 154<br>Purification<br>plant | 9.0-25.0<br>17.0 ± 11.3     | 11.0-29.0<br>20.0 ± 12.7   | 0.8-0.8<br>$0.8 \pm 0.02$ | 18.0-22.3<br>20.4 ± 1.3     | 7.2-16.9<br>11.4 ± 3.2     | 7.4-13.5<br>9.9 ± 1.9     | 7.2-7.6<br>$7.4 \pm 0.2$ | 5.8-8.3<br>7.4 ± 1.1    |
| 9. CB5<br>Agua de leones           | 12.0-90.0<br>29.3 ± 29.9    | 7.0-41.0<br>34.3 ± 29.6    | 0.2-2.1<br>1.0. ± 0.6     | 21.6-29.3<br>25.6 ± 2.0     | 9.7-16.2<br>12.7 ± 1.9     | 7.2-13.7<br>9.7 ± 1.5     | -                        | _                       |
| 10. CB32<br>Agua de leones         | 13.0-18.0<br>$15.0 \pm 2.6$ | 14.0-19.0<br>17.0 ± 2.6    | 0.7-1.0<br>$0.8 \pm 0.1$  | _                           | 9.9-16.7<br>12.7 ± 2.0     | 7.4-13.5<br>10.2 ± 1.9    | _                        | _                       |
| 11. DL18<br>San Borja              | 16.0-47.0<br>31.5 ± 21.9    | 25.0-45.0<br>35.0 ± 14.1   | 0.6-1.0<br>$0.8 \pm 0.2$  | 30.2-49.9<br>41.4 ± 6.1     | 9.2-15.1<br>11.5 ± 2.3     | 7.2-13.3<br>$9.3 \pm 2.3$ | 8.8-10.8<br>9.6 ± 0.7    | 6.7-9.9<br>7.8 ± 1.2    |
| 12. DL26<br>San Borja              | $8.0-12.0$ $10.0 \pm 2.8$   | 9.0-20.0<br>14.5 ± 7.7     | 0.6-0.8<br>$0.7 \pm 0.1$  | _                           | 10.8-15.5<br>12.7 ± 1.7    | 7.2-12.4<br>9.5 ± 1.6     | _                        |                         |
| 13. SL25<br>San Luis<br>Ayucán     | 5.0-20.0<br>11.0 ± 6.4      | 4.0-16.0<br>8.5 ± 5.1      | 0.2-1.5<br>$0.9 \pm 0.6$  | 21.0-28.0<br>$23.0 \pm 2.3$ | 9.9-19.6<br>13.6 ± 2.8     | 8.3-14.9<br>10.0 ± 1.5    | -<br>-                   | <br>-                   |
| 14. VC22<br>Villa del Carbón       | 8.0-29.0<br>15.2 ± 8.3      | 4.0-29.0<br>14.8 ± 10.3    | 0.5-2.0<br>1.2 ± 0.5      | 16.2-19.5<br>18.2 ± 1.0     | 9.2-17.6<br>13.7 ± 2.8     | 7.4-12.8<br>10.0 ± 1.6    | -                        |                         |
| 15. PA3292<br>River Malila         | 19.0-26.0<br>21.4 ± 4.4     | 11.0-28.0<br>21.0 ± 7.1    | 0.6-2.3<br>$0.7 \pm 0.3$  | 20.7-28.0<br>$23.9 \pm 1.9$ | 11.8-19.8<br>14.5 ± 2.7    | 8.6-15.5<br>11.7 ± 1.8    | 17.2-39.5<br>22.9 ± 9.2  | 15.0-25.8<br>18.0 ± 4.4 |
| 16. PA3800<br>River Malila         | 14.0-28.0<br>20.2 ± 5.5     | 9.0-25.0<br>15.6 ± 5.8     | 0.6-2.2<br>$1.4 \pm 0.6$  | _                           | 9.0-19.6<br>14.1 ± 2.8     | 8.8-18.7<br>12.8 ± 2.3    | -                        |                         |
| 17. PA4097<br>River Malila         | $17.0-31.0$ $23.6 \pm 5.5$  | 9.0-19.0<br>14.6 ± 5.1     | $1.0-3.0$ $1.8 \pm 0.8$   | _                           | $10.8-22.3$ $13.9 \pm 2.8$ | 8.3-14.6<br>10.6 ± 1.6    | -                        | _                       |

The second group (Table 1, Figs 11-20) included the three populations from the Malila River, and were identified as P. nevadensis according to the characteristics described by Setchell & Gardner (1920), with some differences in cell size (Table 2). This group was characterized by its small lamina ( $\geq 3.0$  cm long),

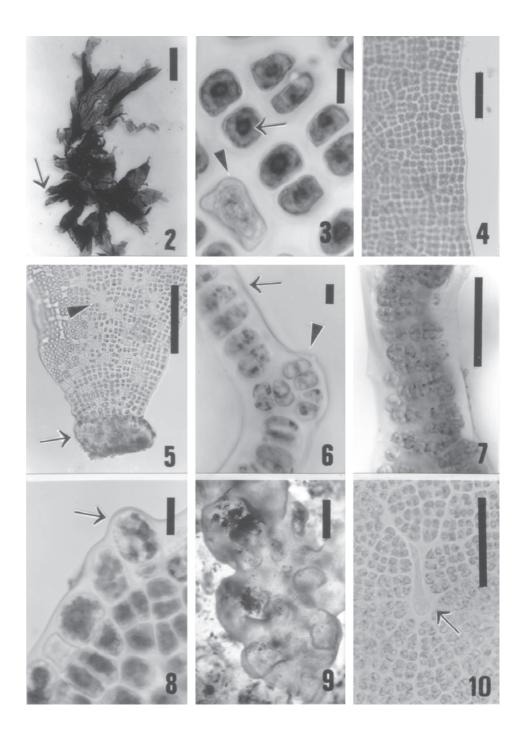


Table 2. Characteristics of freshwater populations of *Prasiola* in Mexico and species reported in the literature.

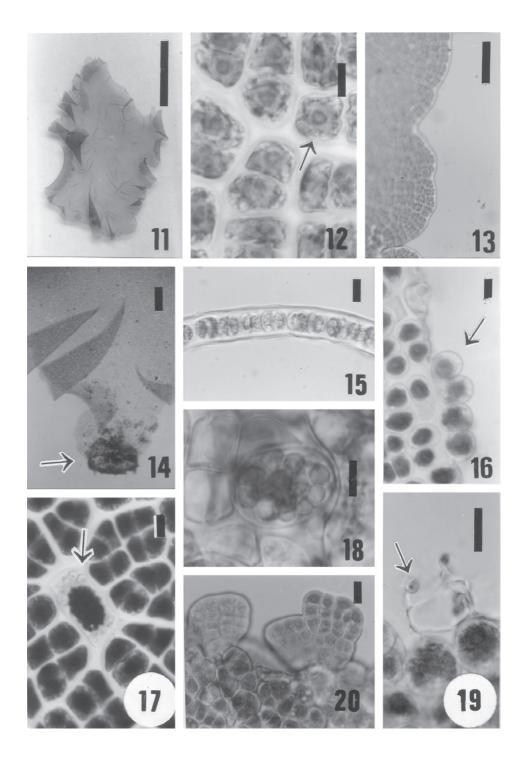
| Taxon                       | Size of frond Width $\times$ length (cm)     | Thickness<br>(µm) | Cell size<br>Width × length<br>(µm) | Reference                                    |
|-----------------------------|--|-------------------|-------------------------------------|--|
| P. mexicana                 | $0.3-4.5 \times 0.2-9$                       | 16.2-130          | $3.1-16 \times 6.1-19.8$            | (this work)                                  |
| P. nevadensis               | $1.4 - 2.1 \times 2 - 2.3$                   | 20.7-28           | $8.3 - 18.7 \times 9 - 22.3$        | (this work)                                  |
| P. fluviatilis              | $0.02 \text{-} 0.3 \times 4.5 \text{-} 95.0$ | 2-4               | $3-5.7 \times 3.2-6.1$              | Hamilton & Edlund (1994)                     |
| P. formosana var. coreana   | $0.4 - 2.3 \times 1.0 - 7.5$                 | 20.0-28.0         | $3-5 \times 10-17.5$                | Naw & Hara (2002)                            |
| P. formosana var. formosana | $a = 0.2 - 2.6 \times 2.0 - 10.7$            | 25.0-30.0         | $2-5 \times 3-7$                    | Naw & Hara (2002)                            |
| P. japonica                 | $0.5 - 4.0 \times 1.0 - 20$                  | 19.0-25.0         | $7.5-10 \times 17.5-22.5$           | Starmach (1972)<br>Naw & Hara (2002)         |
| P. mexicana                 | -× 10.0                                      | 24.0-52.0         | 6.0-16 × –                          | Setchell & Gardner (1920)<br>Starmach (1972) |
| P. nevadensis               | $0.5\text{-}1.0 \times 2.0\text{-}3.0$       | 14.0-25.0         | $2.7 - 4.1 \times 4.1 - 8$          | Setchell & Gardner (1920)                    |
| P. sinica                   | $3.0 \times 9.0$                             | 30.0-33.0         | $5-8 \times 9-13$                   | Starmach (1972)                              |
| P. subareolata              | $0.5 \times 6.0$                             | 45.0-54.0         | $6-8 \times 28-38$                  | Starmach (1972)                              |
| P. velutina                 | $0.03-0.07 (0.11) \times 1-2$                | _                 | -×4-12                              | Starmach (1972)                              |
| P. yunnanica                | $4.0 \times 20.0$                            | 51.0-87.0         | $5.0-7.0 \times 7.0-12.0$           | Starmach (1972)                              |
| Prasiola sp.                | $0.5 \text{-} 5.0 \times 2.0 \text{-} 12.0$  | 20.0-35.0         | $4.0-13.0 \times 12.0-20.0$         | Naw & Hara (2002)                            |

<sup>-</sup> = not reported.

smooth surface, oval form, and one or two fronds that originate from the same base (Fig. 11). It attaches to the substratum with a small mass of elongated, fibrous cells that form a rhizoidal holdfast (Fig. 14). All sections of the lamina tended to be monostromatic and were at least 28  $\mu$ m thick (Fig. 15). Vegetative cells were oval to square, larger than in populations of *P. mexicana*, 8.3-18.7  $\mu$ m in diameter (average 11.7  $\mu$ m); length 9.0-22.3  $\mu$ m (average 14.1  $\mu$ m). In natural conditions round to irregular aplanospores were found with a granular cellular content and dimensions of 15.0-25.8  $\mu$ m in diameter (average 18.0  $\mu$ m) and 17.2-39.5  $\mu$ m in length (average 22.9  $\mu$ m); these were seen to be grouped or isolated at the apex and margin of the lamina surface (Figs 16-17).

Three previously unreported characteristics were observed in the three populations. The margin of the entire lamina was lobed and crenate (Fig. 13). This feature was consistent and allowed the differentiation of *P. nevandensis* from *P. mexicana*. Setchell & Gardner (1920) included figures of *P. nevadensis* where a

Figs 2-10. Morphological characteristics of *Prasiola mexicana*. **2.** General view of the lamina showing bifurcated branches (arrow)(**CB** 5). **3.** Detail of the lobed chloroplast, pyrenoid stained with iodine (arrow), and aplanospores (arrowhead) (specimens in culture). **4.** Detail of the lamina margin (specimens in culture). **5.** Detail of the rhizoidal holdfast (arrow) and a group of aplanospores (arrowhead) (**MA** 26). **6.** Transverse section of the monostromatic region (arrow) and formation of aplanospores (arrow) (**MA** 87). **7.** Transverse section of the polystromatic region (**MA** 87). **8.** Lamina margin with aplanospores (arrow) (specimens in culture). **9.** Surface view of the corrugated lamina (**MA** 87). **10.** Rhizoidal cells in the middle region of the lamina (arrow) (**MA** 87). Scale bars: 10 mm for Figs 2, 9; 100 μm for Figs 4-5, 7, 10; and 10 μm for 3, 6, 8.



lightly crenate margin can be seen, although it was not described in the text: this characteristic, however, is not mentioned for any other freshwater species (Starmach, 1972). The second novelty was the presence of multilocular, ovoid gametangia in the margins of the thallus; these were found to have hyaline gametes with a small pyrenoid, lacking flagella (Figs 18-19), and were from 3.2-5.4 µm in diameter (average 4.3). The gametes were significantly smaller than the aplanospores (p < 0.0001) and agree with the description of the female gametes described for marine populations of *P. stipitata* (3.2-5.0 µm in diameter; Friedmann, 1959a, 1959b) and P. meridionalis (3.0-6.0 µm in diameter; Bravo, 1965). The gametes were liberated upon rupture of the wall of the gametangium (Fig. 19). This is the first report of gametes for this species (Setchell & Gardner, 1920; Starmach, 1979). The third characteristic of the cultured specimens was the development of packets of cells fixed to the apical margin of the adult lamina (Fig. 20); these packets were probably produced by in situ germination of the aplanospores, as described in *P. stipitata* (Friedman, 1959). The cellular packets were found separately and showed a tendency to be filamentous. In order to identify the range of morphological plasticity in this alga, more detailed studies on its reproduction and seasonal morphological changes are needed.

#### **Distribution**

The populations studied were found in 8 waterways of first and second order, at altitudes above 1300 m and in humid to subhumid temperate climates. *Prasiola mexicana* was collected in high montane rivers (altitude > 2,200 m) in areas similar or close to previous records for central Mexico (Setchell & Gardner, 1920; Ramírez *et al.*, 2001). Its distribution is reported to include the United States, Mexico and Costa Rica (Collins 1909; Sherwood *et al.*, 2000; Ramírez *et al.*, 2001). Collections of *P. nevandensis*, previously undescribed in Mexico, represent the second record of the species, and extend its range to the equator (40° N vs. 20° N). On the basis of this record, it is suggested that the distribution of this species might be much broader, as is the case for other temperate North American species found in central Mexico, *Sirodotia suecica* Kylin and *Paralemanea annulata* (Kützing) Vis *et* Sheath (Carmona, 1997; Carmona & Necchi, 2002). It will be necessary to explore waterways of similar characteristics (mountainous areas with cold water) in more tropical latitudes to confirm this hypothesis.

In terms of environmental characteristics (Table 3), populations of *P. mexicana* grew in seven shallow (0.1-20.0 cm) rivers with cold water (2.0°C a 16.4°C), a slightly acidic to basic pH (6.4-7.6), low conductivity (35.5-94.2 μS.cm<sup>-1</sup>),

Figs 11-20. Morphological characteristics of *Prasiola nevadensis*. **11.** General view of the lamina (**PA** 4092). **12.** Detail of the lobed chloroplast and pyrenoid stained with iodine (arrow) (specimens in culture). **13.** Detail of the lamina margin (**PA** 4092). **14.** Detail of the rhizoidal holdfast (arrow) (**PA** 3800). **15.** Transverse section of the monostromatic region (**PA** 4092). **16.** Detail of the lamina margin with a group of aplanospores (arrow) (specimens in culture). **17.** Aplanospores isolated with granular cytoplasm (arrow) (specimens in culture). **18.** Detail of the gametangium in the lamina margin (specimens in culture). **19.** Liberation of gametes (arrow) via rupture of the gametangial wall (specimens in culture). **20.** Cellular packets fixed to the apical margin of the adult lamina (specimens in culture). Scale bars: 10 mm for Figs 11, 15; 100 μm for Figs 13-14; and 10 μm for 12, 16-20.

Table 3. Physical and chemical characteristics of streams containing freshwater populations of *Prasiola* in Central Mexico. Number of sampling sites are those represented in Figure 1.

| Sampling site                                | Populations   | Altitude<br>(masl) | Climate <sup>a</sup> | Aliiude Climate <sup>a</sup> Temperature (masl) | Hd      | Specific<br>conductance<br>(μS.cm <sup>-1</sup> ) | Current velocity (cm.s <sup>-1</sup> ) | Current Shading <sup>b</sup> Depth velocity (cm) | Depth<br>(cm) | Substratum <sup>c</sup> | Associated species  |
|--|---------------|--------------------|----------------------|---|---------|---|--|--|---------------|-------------------------|---|
| 1. Trout tank,<br>river La Magdalena         | -             | 3000               | $C(w_2)(w)$          | 4   | 6.4     | 65  | 09                                     | A-C  | 0-20          | $\vdash$                | Cocconeis placentula var. lineata<br>(Ehrenberg) Van Heurck, Hydrococcus<br>cesatii, Rabenhorst, Cymbella silesiaca<br>Bleisch.   |
| 2. Xalancocotta,<br>river La Magdalena       | 2, 3, 4, 5, 6 | 3150               | s                    | 3-11  | 6.7-7.7 | 06-59   | 0-85                                   | A-C  | 6             | 1-2                     | Synedra ulna (Nitzch) Ehrenberg Metosira varians I. Agardh, Cymbella mexicana (Ehrenberg) Cleve, Gomphonema sp., Diploneis elliptica (Kützing) Cleve, Placoma regulare Broady et Ingerfeld. |
| 3. Purification plant,<br>river La Magdalena | 7,8           | 2800               | S                    | 2-11  | 7.3-7.6 | 90-94   | 0-85                                   | C  | 0-11          | 1-2                     | Ulothrix sp.  |
| 4. Agua de Leones                            | 9, 10         | 3220               | s                    | 7-12  | 7.0-7.6 | 35-36   | 06                                     | A  | 0-5           | $\vdash$                | C. placentula var. lineata, Xenotholos sp., Fragilaria sp., P. regulare   |
| 5. San Borja                                 | 11, 12        | 2800               | s                    | 10-16   | 6.9-7.2 | 09-09   | 35-85                                  | Ą  | 0-10          | 3                       | C. placentula var. lineata, Cymbella sp.,<br>Navicula sp.   |
| 6. San Luis Ayucán                           | 13            | 2664               | s                    | 16  | 9.9     | 79  | 09                                     | Y  | 10            | s                       | Paralemanea mexicana (Kützing)<br>Vis et Sheath   |
| 7. Villa del Carbón                          | 14            | 2227               | s                    | 14  | 7.0     | 92  | 35-85                                  | Ą  | 8-0           | S                       | I   |
| 8. River Malila                              | 15, 16, 17    | 1300               | C(fm)                | 14-17   | 5.5-7.2 | 192   | 60-85                                  | O  | 0-20          | S                       | C. placentula var. euglypta (Ehrenberg) Grunow, Xenotholos huastecanus Gold, Montejano et Komárek, Chamaesiphon sp.   |

 $^a$  = García (1973): C(w<sub>2</sub>)(w) = Temperate, dry winter < 5%; C (fm) = Temperate, no dry season.  $^b$  = A, open; B, parttly shaded; C, shaded.  $^c$  = 1, predominantly bedrock; 2, predominantly boulder.

and a wide range of light intensity (low-high) and water current speed (10-85 cm.s<sup>-1</sup>). In populations 2 and 10, specimens were collected in the splash zone from the water to 10 cm above the maximum level of the current. The temperature data are similar to those reported by Sheath & Cole (1992) and Ramírez *et al.* (2001). The most frequent substratum was andesitic-basaltic rock. Occasionally trunks or metal tubes served as a substratum as well. *Prasiola mexicana* commonly grew with other algae such as *Placoma regulare* and *Cocconeis placentula* var. *lineata*, and was occasionally seen with *Paralemanea mexicana*. Populations of *P. nevadensis* grew in a river of humid temperate climate with cold water (14.0°C) that was acidic (pH = 5.5), had medium specific conductivity (192 μS.cm<sup>-1</sup>), was shallow (0.2-20.0 cm), and had low irradiance and high water current speed (60-85 cm.s<sup>-1</sup>). The predominant substratum was lime-lutite and *P. nevadensis* grew in association with other algae such as *Xenotholos huastecanus*. This is the first report of the habitat of *P. nevadensis*.

# **Description of the Mexican material**

# Prasiola mexicana J. Agardh, 1847. (Figs 2-10)

Monostromatic thallus 2.0-9.0 cm long, 0.3-4.5 cm wide, length/width 0.2-3.7; with bifurcating ramifications at apex. Lamina surface smooth or corrugated and margin smooth to lobed; thickness of 16.2-130.0  $\mu$ m; 1 to 4 laminae per rhizoidal hold-fast. Cells oval to rectangular, 6.1-19.8  $\mu$ m long, 3.1-16.0  $\mu$ m in diameter. Aplanospores 4.5-10.8  $\mu$ m long, 5.4-10.6  $\mu$ m in diameter in the polystromatic layer (4-8 layers).

Populations examined: (Numbering corresponds to the list of Table 1). 1) D.F., Magdalena Contreras, Trout tank, coll. J. Carmona, 19.xii.1998 (FCME MA87); 2) Xalancocotla, coll. J. Carmona, 23.xi.1997 (FCME MA1); 3) coll. M. Ramírez, 23.xi.1997 (FCME MA9); 4) coll. J. Carmona, 8.ii.1998 (FCME MA26); 5) (FCME MA36); 6) coll. M. Bojorge, 4.xi.1999 (FCME MA102); 7) Purification plant, coll. J. Carmona, 24.i.1999 (FCME MA94); 8) coll. M. Ramírez, (FCME MA154); 9) Cuajimalpa, Agua de Leones, coll. J. Carmona, 19.vi.1999 (FCME CB5); 10) coll. E. Cantoral, 28.xi.1999 (FCME CB32); 11) San Borja, coll. J. Carmona, 15.v.1999 (FCME DL18); 12) coll. G. Montejano, 15.v.1999 (FCME DL26); 13) Estado de México, Jilotzingo, San Luis Ayucán, coll. M. Ramírez, 18.vii.2001 (FCME SL25); 14) Villa del Carbón, river Villa del Carbón, coll. J. Carmona, 18.vii.2001 (FCME VC22).

## **Prasiola nevadensis** Setchell et Gardner, 1920. (Figs 11-20)

Monostromatic thallus, 2.2-2.3 cm long, 1.4-2.1 cm wide, length/width 0.5-2.3. Lamina surface smooth, margin lobed and crenate; lamina 20.7-28.0  $\mu m$  thick. 1-2 laminae per rhizoidal holdfast. Cells oval to rectangular, 9.0-22.3  $\mu m$  long, 8.3-18.7  $\mu m$  diameter. Aplanospores 17.2-39.5  $\mu m$  long, 15.0-25.8  $\mu m$  in diameter, in the polystromatic layer (1-2 layers). Gametangia ovoid with elliptical or oval gametes, without flagella, 3.2-5.4  $\mu m$  in diameter.

**Population examined:** (Numbering corresponds to the list of Table 1) **15)** Hidalgo, Molango, river Malila, *coll. J. Carmona*, 15.ii.1992 (FCME PA3292); **16)** *coll. G. Montejano*, 10.iii.1999 (FCME PA3800); **17)** *coll. J. Carmona*, 6.xi.2002 (FCME PA4097).

**Acknowledgements.** This research was supported by a CONACYT (J33930-V) Research Grant to E. Cantoral. We would like to thank Isabel Bieler Antolín and Yenny Beltrán Magos for her help in printing photographs, Laura Patricia Olguín of the controlled environment chambers for their help in maintaining cultures, Jeffrey Ross-Ibarra for reviewing the English and Pierre Compère for French translation.

#### REFERENCES

- AGARDH J., 1847 Nya alger fran Mexico. Öfversigt af Kongl. Vetenskaps-Akademiens Förhandlinglar 4(1): 5-17.
- BRAVO L.M., 1965 Studies on the life history of *Prasiola meridionalis*. *Phycologia* 4: 177-194. CARMONA J.J., 1997 *Estudio florístico (Taxonómico-ecológico-biogeográfico) de las rodofitas de agua dulce en la región central de México*. PhD Thesis, México, DF, Universidad Nacional Autónoma de México, 201 p.
- CARMONA J.J. & NECCHI O.Jr., 2002 Somatic meiosis and development of the juvenile gametophyte in members of the Batrachospermales *sensu lato* (Rhodophyta). *Phycologia* 41: 340-374.
- COLLINS F.S., 1909 The green algae of North America. *Tufts College Studies* 2: 79-490.
- FRIEDL T. & O'KELLY C.J., 2002 Phylogenetic relationship of green algae assigned to the genus *Planophila* (Chlorophyta): evidence from 18S rDNA sequence data and ultrastructure. *European Journal of Phycology* 37: 373-384.
- FRIEDMANN I., 1959a Structure, life-history, and sex determination of *Prasiola stipitata* Suhr. *Annals of Botany* 23: 571-594.
- FRIEDMANN I., 1959b Gametes, fertilization and zygote development in *Prasiola stipitata* Suhr. *Nova Hedwigia* 1: 333-44.
- FRITSCH F.E., 1961 The structure and reproduction of the algae. Vol I Norwich, Jarrold & Sons Ltd, 939 p.
- GARCÍA E., 1973 Modificaciones al sistema de clasificación de Köppen (para adaptarlo a las condiciones de la República Mexicana). México, Instituto de Geografía, Universidad Nacional Autónoma de México, 246 p.
- HAMILTON P.B. & EDLUND S.A., 1994 Occurrence of *Prasiola fluviatilis* (Chlorophyta) on Ellesmere Island in the Canadian Artic. *Journal of Phycology* 30: 217-221.
- HOLMGREN P.K., HOLMGREN N.H. & BARNETT L.C., 1990 *Index Herbariorum.* Part I. The herbaria of the world. 8th edition. New York, New York Botanical Garden, 693 p.
- JOHANSEN S.A., 1940 Plant microtechnique. New York, Mc Graw-Hill, 523 p.
- NAW M.W. & HARA Y., 2002 Morphology and molecular phylogeny of *Prasiola* sp. (Prasiolales, Chlorophyta) from Myanmar. *Phycological Research* 50: 175-182.
- O'KELLY C.J., GARBARY D.J. & FLOYD G.L., 1989 Flagellar apparatus of male gametes and other aspects of gamete and zygote ultrastructure in *Prasiola* and *Rosenviengiella* (Chlorophyta, Prasiolales) from British Columbia. *Canadian Journal of Botany* 67: 505-514.
- ORTEGA M.M., 1984 Catálogo de algas continentales recientes de México. México, DF, Dirección General de Publicaciones, Universidad Nacional Autónoma de México, 566 p.
- RAMÍREZ V.M., BELTRÁN M.Y., BOJORGE G.M., CARMONA J.J., CANTORAL U.E. & VALADEZ F., 2001 Flora algal del Río La Magdalena, Distrito Federal, México. *Boletín de la Sociedad Botánica de México* 68: 51-73.
- RINDI F., GUIRY M.D., BARBIERO R. P. & CINELLI F., 1999 The marine and terrestrial *Prasiolales* (Chlorophyta) of Galway City, Ireland: a morphological and ecological study. *Journal of Phycology* 35: 469-82.
- SETCHELL W.A. & GARDNER N. L., 1920 The marine algae of the Pacific Coast of North America. Part 2. Chlorophyceae. *University of California Publications in Botany* 7: 139-374.
- SHEATH R.G. & COLE K.M., 1992 Biogeography of stream macroalgae in North America. *Journal of Phycology* 28: 448-460.
- SHERWOOD A.R., GARBARY D.J. & SHEATH R.G., 2000 Assessing the phylogenetic position of the *Prasiolales* (Chlorophyta) using rbcL and 18S rRNA gene sequence data. *Phycologia* 39: 139-46.
- SOUTHWOOD T.R.E., 1978 *Ecological methods with particular reference to the study of insect populations*. 2<sup>nd</sup> Ed. New York, Chapman and Hall, 524 p.
- STARMAČH K., 1972 *Flora slodkowodna Polski*. Tômo 10: Chlorophyta III. Warszawa-Kraków, Polska Akademia Nauk, 750 p.
- VAN DEN HOEK C., MANN D.G. & JAHNS H.M., 1995 Algae. An introduction to phycology. Cambridge, University Press, 623 p.